

# Single-step genetic evaluation of resistance to parasites in the Swiss Alpine goat population

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30.8.2021

**EAAP Davos** 

#### Resistance to parasites

- Gastrointestinal nematodes are source of one of the most important disease in small ruminants in pasture-based production system.
- After decades of the use of anthelminic products, parasites developed resistances.
- As alternative to anthelminic product, breeding for more resistant animals was proposed in sheep and goats

#### Previous study

Parasite 2017, **24**, 32

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DOI: 10.1051/parasite/2017033



RESEARCH ARTICLE

OPEN 3 ACCESS

#### The genetic basis for the selection of dairy goats with enhanced resistance to gastrointestinal nematodes

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Received 19 December 2016, Accepted 14 July 2017, Published online 9 August 2017

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#### Phenotypes

- Measures of fecal egg count (FEC), packed cell volume (PCV) and FAMACHA© eye color score (FAA).
  - 948 Alpine goats in 17 Swiss herds
- Higher parasitic load leads to:
  - higher fecal egg count
  - lower percentage of packed cell volume, and
  - whiter FAMACHA© eye color score.
- All traits used together could build a resistance index

#### **Dataset**

- Pedigree contained 5652 animals
  - Of which 1277 were genotyped (60K Goat Chip V2)
- Heritability estimates, genetic (above diag) and phenotypic (below diag) correlations:

	FEC	FAA	PCV
FEC	0.07	-0.03	-0.39
FAA	0.18	0.22	-0.60
PCV	-0.27	-0.17	0.22

#### Phenotyping

- Two measures
  - Early summer (after 3-4 months on pasture)
  - Early autumn (3-4 months after helminthic treatment)
- FEC transformed to obtain normal distribution:
   FEC<sub>t</sub> = (FEC + 1)<sup>0.36</sup>
- FEC reduction test (FECRT) done to account for level of resistance in each herd

#### Multi-traits animal model

 $FEC_t$  = season + anthelmintic + FECRT + age class + herd + pe + a + e

FAA = season + anthelmintic + FECRT + age class + classifier + herd + pe + a + e

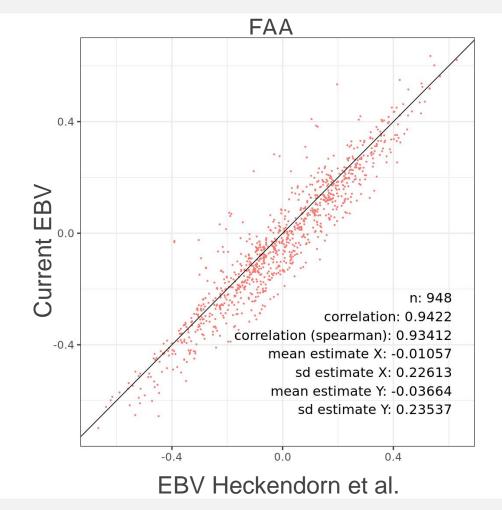
PCV = season + anthelmintic + FECRT + age class + herd + pe + a + e

#### Reproduction of PBLUP by Heckendorn et al.

- Differences expected:
  - Multibreed vs only Alpine
  - Different pedigrees
     2012 vs 2021
  - Different evaluation software aireml90 vs MiX99 suite

#### BUT

- Same variance components
- Same phenotypic dataset



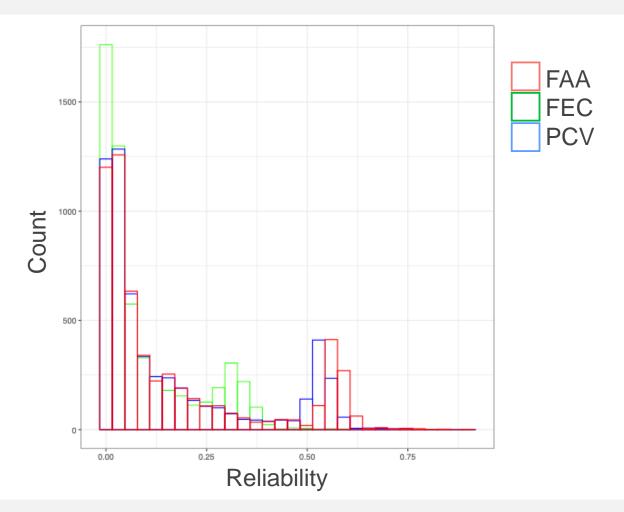
#### Reliabilities of non-genomic breeding values

- Reliability distribution similar for all three traits
- Averages are very low:

• FAA: 0.164

• FEC: 0.096

• PCV: 0.156



#### Reliabilities of non-genomic breeding values

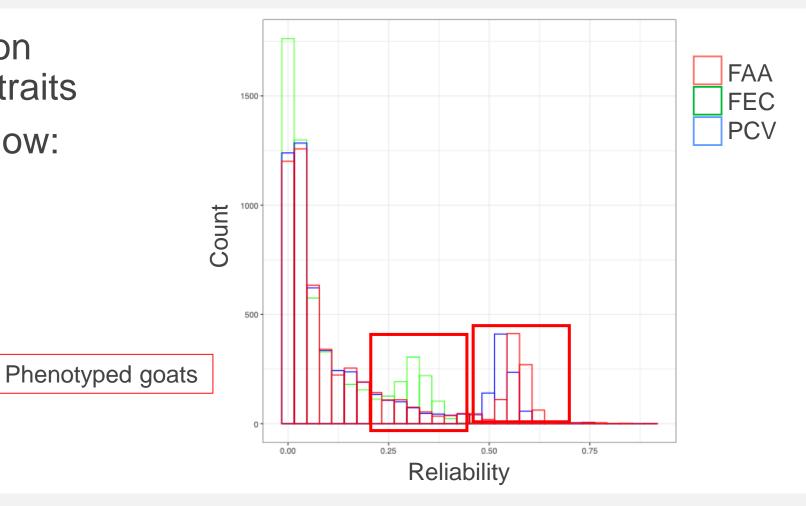
 Reliability distribution similar for all three traits

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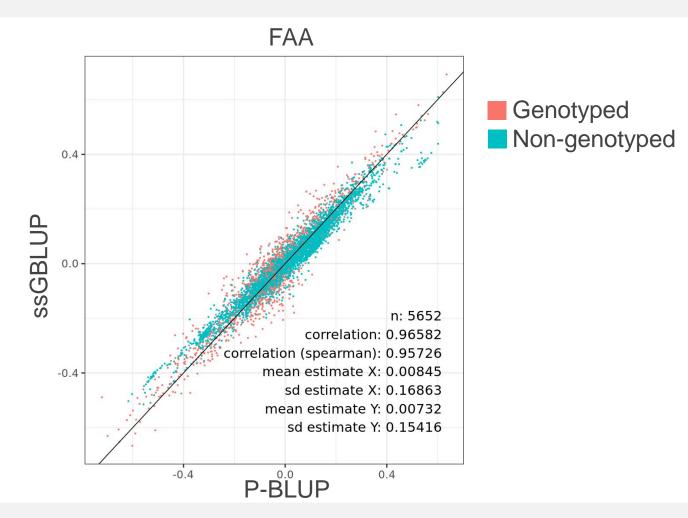
• PCV: 0.156



#### From P-BLUP to ssGBLUP

- 16% of the animals in evaluation were phenotyped and genotyped
- No clear EBV difference between genotyped and nongenotyped animals
- High correlation between PBLUP and ssGBLUP

==> Dataset still very small for traits with low heritability estimates



- Use pipeline differentiating genotyped and non-genotyped animals:
  - Estimate reliabilities for all animals without genomic information (P-BLUP)
  - Estimate genomic reliabilities for genotyped animals (SNP-BLUP)
  - Integrate additional genomic information into P-BLUP model
  - Estimate genomic reliabilities for non-genotyped animals

#### Approximate individual animal reliabilities in single-step genomic model

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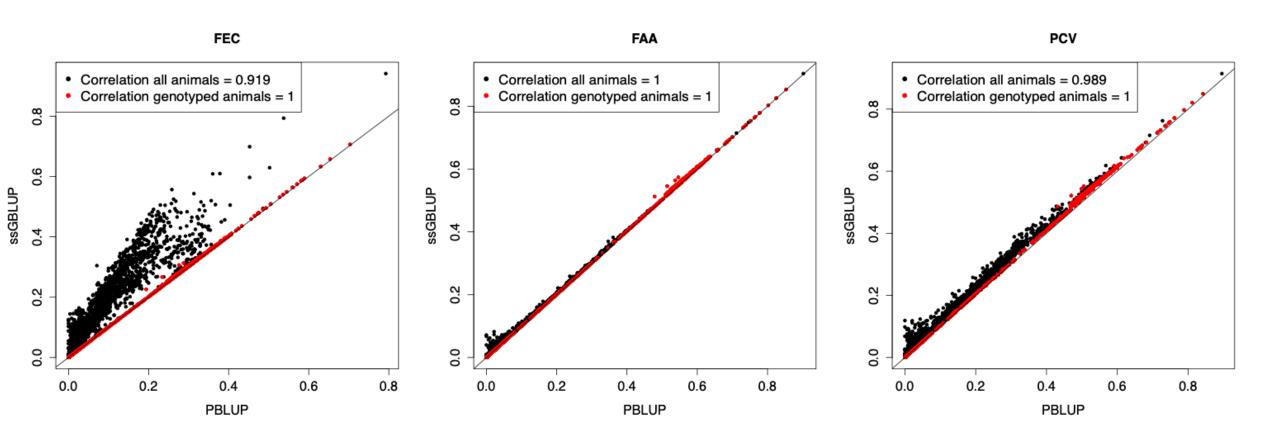
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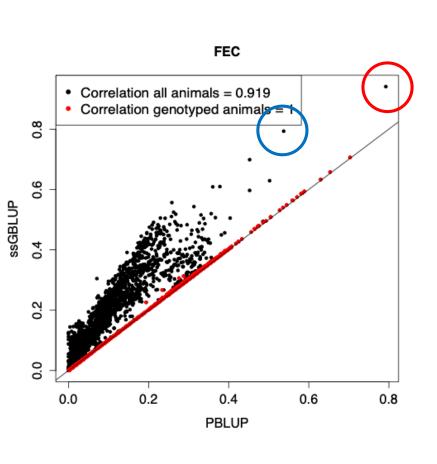
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Buck of 87 animals in pedigree, of which 79 are genotyped.

Buck of 25 animals in pedigree, of which 17 are genotyped.

#### Conclusions

- Reproduction of the work by Heckendorn et al. was possible with MiX99 and additional pedigree information
  - EBV correlations of 0.94
  - Reliability estimates were still very low
- Change from PBLUP to ssGBLUP approach did not show any great impact on EBV for any genotyped or non-genotyped animals
- Reliability estimates are increased with additional genomic information, but data must be further expanded before any possible implementation.

#### Acknowledgements

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## Thank you for your attention



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